



# Antibacterial and Antioxidant Properties of Methanolic Extracts of Some Native Edible Plants Collected from Kermanshah, Western Iran

Y. Shahbazi \*✉

Department of Food Hygiene and Quality Control, Faculty of Veterinary Medicine, Razi University, Kermanshah, Iran

## HIGHLIGHTS

- The most antibacterial effectiveness was found for *Mentha longifolia* extract.
- Gram-negative bacteria were more resistant to the presence of methanolic plant extracts than Gram-positive bacteria.
- The highest antioxidant activity was found in *M. longifolia* extract.
- *Falcaria vulgaris*, *Allium rotundum*, *Tragopogon graminifolius*, and *M. longifolia* can be used for food preservation.

## Article type

Original article

## Keywords

Anti-Bacterial Agents  
Antioxidants  
Plants

## Article history

Received: 13 Aug 2017  
Revised: 2 Nov 2017  
Accepted: 27 Nov 2017

## Acronyms and abbreviations

BHT=Butylated Hydroxytoluene  
BHI=Brain Heart Infusion  
MIC=Minimum Inhibitory Concentration  
MBC=Minimum Bactericidal Concentration  
CFU=Colony Forming Unit  
DPPH=2, 2-diphenyl-1-picrylhydrazyl hydrate

## ABSTRACT

**Background:** There is growing demand to improve physicochemical, microbiological, and sensory properties of fresh foods using natural herbal antimicrobial and antioxidant compounds. The aim of the present study was to investigate antioxidant and antibacterial properties of some native edible plants of Kermanshah, Western Iran.

**Methods:** The methanolic extracts of leaves of *Falcaria vulgaris*, *Allium rotundum*, *Tragopogon graminifolius*, and *Mentha longifolia* plants were prepared. The antibacterial effects of these four plant extracts were determined on *Salmonella typhimurium*, *Bacillus subtilis*, *B. cereus*, *Staphylococcus aureus*, *Listeria monocytogenes*, and *Escherichia coli* O157:H7 using micro-broth dilution and agar disk diffusion assays. Also, the 2, 2-diphenyl-1-picrylhydrazyl hydrate assay was used for determination of antioxidant properties of the plant extracts. The analysis was performed using SPSS 16.0 (Chicago, IL, USA) software package.

**Results:** The most antibacterial effectiveness was significantly ( $p<0.05$ ) found for *M. longifolia* extract. The following sequence inhibition effect on investigated bacterial strains was observed: *M. longifolia*>*T. graminifolius*>*A. rotundum*>*F. vulgaris*. Moreover, Gram-negative bacteria were more resistant to the presence of methanolic plant extracts than Gram-positive bacteria. The highest antioxidant activity (based on IC<sub>50</sub>) was significantly ( $p<0.05$ ) found for *M. longifolia* ( $0.88\pm 0.12$  mg/ml); as well as these rates for *T. graminifolius*, *A. rotundum*, and *F. vulgaris* extracts were  $0.45\pm 0.78$ ,  $0.26\pm 0.07$ , and  $0.14\pm 0.23$  mg/ml, respectively.

**Conclusion:** The studied edible plants had antimicrobial and antioxidant activities that recommend as potential preservatives in food products. However, methanolic extract of *M. longifolia* had the best antibacterial and antioxidant properties *in vitro*.

## Introduction

It has been reported that garlic, onion, basil, cinnamon, curry, mustard, ginger, and also other spices exhibit anti-

microbial and antioxidant properties (Alzoreky and Nakahara, 2003; Benkeblia, 2004; Hussain et al., 2008; Park et al., 2008; Sakunpak and Panichayupakaranant, 2012). Also, antimicrobial and antioxidant compounds derived from natural sources like plants have been tradi-

\*Correspondence. ✉ y.shahbazi@razi.ac.ir

**To cite:** Shahbazi Y. (2017). Antibacterial and antioxidant properties of methanolic extracts of some native edible plants collected from Kermanshah, Western Iran. *Journal of Food Quality and Hazards Control*. 4: 93-98.

tionally used for treatment of various diseases especially cancers as well as gastroenteritis (Runyoro et al., 2010). Epidemiological studies have indicated that there is a remarkable positive association between the high intake of fresh fruits and vegetables and a reduction rate of heart disease, mortality, different types of cancers, and other degenerative diseases (Conforti et al., 2009; Pal et al., 2012; Schinella et al., 2002; Shahbazi et al., 2015; Yang et al., 2016).

Food-borne diseases have serious negative effects on public health and food security (Shahbazi and Shavisi, 2016). Moreover, there is growing demand to improve physicochemical, microbiological, and sensory properties of fresh foods using natural herbal antimicrobial and antioxidant compounds (Shahbazi et al., 2016). The use of antioxidants in food owing to their ability to control fat oxidation as well as increase the shelf life of the food is highly appreciated in food industries. butylated hydroxyanisole and Butylated Hydroxytoluene (BHT) are the most common synthetic antioxidants which is used in food products (Shahbazi et al., 2015). Despite the effectiveness and good stability, the use of these compounds due to their toxicity is limited in modern preservation methods (Alzoreky and Nakahara, 2003). Therefore, a particular tendency to use a healthy compounds such as edible plants and vegetables in food has arisen (Pal et al., 2012). Because of strong antioxidant and antimicrobial activities which are related to high levels of flavonoids and phenolic acid, some edible plants and vegetables are recommended as potential additives for food preservation (Sakunpak and Panichayupakaranant, 2012).

Previous studies indicated that extracts of various edible plants have strong antibacterial and antioxidant activities in food models as well as *in vitro* (Asekun et al., 2007; Gulluce et al., 2007; Jaberian et al., 2013; Mkaddem et al., 2009; Monfared et al., 2012; Shafaghat, 2010). However, a study on the antibacterial and antioxidant activities of *Falcaria vulgaris* (Paghazeh), *Allium rotundum* (Sir-e-Kouhi), *Tragopogon graminifolius* (Sheng), and *Meniha longifolia* (Pouneh) collected from Kermanshah, West of Iran has not been carried out so far. Therefore, the aim of the present study was to investigate antioxidant and antibacterial properties of methanolic extracts of some native edible plants of Kermanshah, Western Iran.

## Materials and methods

### Collection of plant materials

The fresh leaves of *F. vulgaris*, *A. rotundum*, *T. graminifolius*, and *M. longifolia* plants were collected from Kermanshah, West of Iran during full flowering

period (March to April 2016). The fresh leaves were air-dried in a shadow at  $25 \pm 1$  °C for 14 days. Authentications of the plants were done by Faculty of Agriculture, Razi University, Kermanshah, Iran. The voucher numbers of *F. vulgaris*, *A. rotundum*, *T. graminifolius*, and *M. longifolia* plants were 7163, 87981, 9160, and 3289, respectively.

### Preparation of plant extracts

Five g of each fine-powdered plant was dissolved in 20 ml methanol and extracted with a shaker at  $25 \pm 1$  °C for 24 h. The extract was then filtered by Whatman filter paper no. 3, concentrated in a rotary evaporator (IKA, Germany) at 40 °C and stored at refrigerator ( $4 \pm 1$  °C) till next analysis (Motamed and Naghibi, 2010).

### Preparation of bacterial strains

*Salmonella typhimurium* (ATCC 14028), *Bacillus subtilis* (ATCC 6633), *B. cereus* (ATCC 11774), *Staphylococcus aureus* (ATCC 6538), *Listeria monocytogenes* (ATCC 19118), and *Escherichia coli* O157:H7 (ATCC 10536) were obtained from the culture collection of the Iranian Research Organization for Science and Technology, Tehran, Iran. The standard bacterial strains were incubated at 37 °C for 24 h after inoculation in Brain Heart Infusion (BHI) broth medium, adjusted to a final density of  $10^5$  Colony Forming Unit (CFU)/ml as an inoculum dose (Carović-Stanko et al., 2010).

### Determination of the Minimum Inhibitory Concentration (MIC) and the Minimum Bactericidal Concentration (MBC)

The MIC values of methanolic plant extracts were determined using micro-broth dilution assay. Different concentrations of extracts ranging from 0.5 to 10 mg/ml were set up by serial dilution in BHI (Merck, Darmstadt, Germany) broth containing dimethyl sulfoxide (0.5% v/v). The 96-well sterile micro-titer plates were prepared by dispensing 180 µl BHI broth medium containing specified concentrations of the antibacterial agents and 20 µl bacterial inoculum ( $5 \log$  CFU/ml) into each well. BHI broth containing inoculum without the analyzed materials (as positive control) and BHI broth containing the analyzed materials (as negative control) were evaluated in the last wells of each strip. The microplates were shaken at 300 rpm for 20 s and incubated at 37 °C for 24 h. The lowest concentration that completely inhibited the growth of microorganisms was defined as MIC value. To determine MBC, 20 µl of the content of each well with no invisible growth was sub-cultured on BHI (Merck, Darmstadt, Germany) agar and incubated at 37 °C for 24 h. The concentration of each extract in those wells that

yielded plates with no visible colonies was considered to be the MBC (Alves-Silva et al., 2013; Carović-Stanko et al., 2010; Shahbazi and Shavisi, 2016).

#### Agar disk diffusion assay

For agar disk diffusion assay, 0.1 ml of each bacterial suspension ( $10^8$  CFU/ml) was uniformly spread on BHI agar medium using a sterile cotton swab. Then, the sterile paper disks (6 mm in diameter) incorporated with 10  $\mu$ l of the highest concentration of each plant extracts (10 mg/ml) was placed on the surface of each BHI agar. The plates were incubated for 24 h at 37 °C. In order to evaluate antibacterial activity, the diameter of inhibition zone was measured, then disk diameter (6 mm) was deducted and the inhibition zone was reported as  $\pi r^2$  ( $\pi=3.14$  and  $r$ =radius of inhibition zone which described previously by Ghasemi Pirbalouti et al. (2016).

#### Antioxidant analysis of methanolic plant extracts

The 2, 2-diphenyl-1-picrylhydrazyl hydrate (DPPH) assay was used for determination of antioxidant properties of plant extracts. Stock solutions (100 mg/ml) of each plant extract as well as BHT (Merck, Darmstadt, Germany) as the synthetic standard antioxidant were prepared in methanol. Selected diluted concentrations were mixed to 1 ml DPPH methanol solution. The mixtures were then shaken and allowed to stand for about 30 min at  $25\pm 1$  °C in the dark. Tests were conducted for investigated extracts and BHT (as synthetic antioxidant). The ultraviolet absorbencies of these solutions were measured at 517 nm using visible light spectrophotometer. Finally, the DPPH radical scavenging activity was evaluated as follow (Wannes et al., 2010):

$$\text{DPPH (I\%)} = [(A_{\text{blank}} - A_{\text{sample}}) / A_{\text{blank}}] \times 100$$

Where  $A_{\text{blank}}$  is the absorbance of the blank sample and  $A_{\text{sample}}$  is the absorbance of the plant extract. The sample concentration providing 50% inhibition ( $IC_{50}$ ) was calculated from the curve of radical scavenge activity (I%) against sample concentration.

#### Statistical analysis

All experiments were carried out in triplicate. The analysis was performed using SPSS 16.0 for Windows (Chicago, IL, USA) software package. The significance level was considered  $p < 0.05$  in all experimental data obtained in this research.

## Results

The antibacterial effects of *F. vulgaris*, *A. rotundum*, *T. graminifolius*, and *M. longifolia* extracts against common

food-related pathogens, including *S. aureus*, *B. subtilis*, *B. cereus*, *L. monocytogenes*, *S. typhimurium*, and *E. coli* O157:H7 are exhibited in Tables 1 and 2. As it can be seen, the most antibacterial effectiveness was significantly ( $p < 0.05$ ) found for *M. longifolia* extract, which inhibition zone and MIC were in the range of 3.14-12.45 mm and 2-10 mg/ml, respectively. The following sequence inhibition effect on investigated bacterial strains was observed: *M. longifolia* > *T. graminifolius* > *A. rotundum* > *F. vulgaris*.

Moreover, Gram-negative bacteria (*S. typhimurium* and *E. coli* O157:H7) were more resistant to the presence of methanolic plant extracts than Gram-positive bacteria (*S. aureus*, *B. subtilis*, *B. cereus*, and *L. monocytogenes*). *B. cereus* was significantly ( $p < 0.05$ ) the most sensitive bacterium for *M. longifolia*, *T. graminifolius*, *A. rotundum*, and *F. vulgaris* extracts with inhibition zones of  $12.45\pm 0.03$ ,  $10.56\pm 0.01$ ,  $7.04\pm 0.01$ , and  $4.24\pm 0.01$  mm, respectively.

Based on the results of the present study, the highest  $IC_{50}$  was significantly ( $p < 0.05$ ) found in *M. longifolia* ( $0.88\pm 0.12$  mg/ml); these rates for *T. graminifolius*, *A. rotundum*, and *F. vulgaris* extracts were  $0.45\pm 0.78$ ,  $0.26\pm 0.07$ , and  $0.14\pm 0.23$  mg/ml, respectively.

## Discussion

In general, it is assumed that the considerable antibacterial activities of *F. vulgaris*, *A. rotundum*, *T. graminifolius*, and *M. longifolia* extracts could be related to the phenolic compounds; the hydroxyl groups in phenolic compounds are thought to lead inhibitory action as these groups can interact with bacterial cell membrane to disrupt membrane structures and cause the leakage of cellular components. This will lead to the collapse of the proton motive force and depletion of the ATP pool and ultimately resulting in cell death (Gyawali and Ibrahim, 2014; Jay et al., 2005). On the other hand, the significant difference in the antimicrobial action of plant extracts is probably due to variability in the nature and concentration of main groups of chemical compositions (Lv et al., 2011). Moreover, we found that Gram-negative bacteria were more resistant to the presence of investigated methanolic plant extracts than Gram-positive bacteria. It may be related to the hydrophobic outer membrane surrounding the bacterial cell wall that restricts diffusion of lipophilic materials like extracts (Shahbazi and Shavisi, 2016). Some similar investigations on the *in vitro* antibacterial activities of *F. vulgaris*, *A. rotundum*, *T. graminifolius*, and *M. longifolia* extracts have been conducted (Alzoreky and Nakahara, 2003; Sakunpak and Panichayupakaranant, 2012; Shafaghat, 2010). Jaberian et al. (2013) collected *F. vulgaris* from

**Table 1:** Antibacterial effect of methanolic extracts of *Falcaria vulgaris*, *Allium rotundum*, *Tragopogon graminifolius*, and *Mentha longifolia* indicated as Minimum Inhibitory/Bactericidal Concentrations (MIC/MBC; mg/ml)

| Bacteria                | <i>M. longifolia</i> |                     | <i>T. graminifolius</i> |                    | <i>A. rotundum</i> |                     | <i>F. vulgaris</i> |                     |
|-------------------------|----------------------|---------------------|-------------------------|--------------------|--------------------|---------------------|--------------------|---------------------|
|                         | MIC                  | MBC                 | MIC                     | MBC                | MIC                | MBC                 | MIC                | MBC                 |
| <i>S. aureus</i>        | 4±0.0 <sup>b</sup>   | 4±0.0 <sup>b</sup>  | 8±0.0 <sup>a</sup>      | 8±0.0 <sup>a</sup> | 8±0.0 <sup>a</sup> | 10±0.0 <sup>a</sup> | >10 <sup>a</sup>   | >10 <sup>a</sup>    |
| <i>B. subtilis</i>      | 2±0.0 <sup>b</sup>   | 4±0.0 <sup>b</sup>  | 6±0.0 <sup>b</sup>      | 6±0.0 <sup>b</sup> | 8±0.0 <sup>a</sup> | 8±0.0 <sup>a</sup>  | 8±0.0 <sup>a</sup> | 10±0.0 <sup>a</sup> |
| <i>B. cereus</i>        | 2±0.0 <sup>b</sup>   | 2±0.0 <sup>b</sup>  | 6±0.0 <sup>b</sup>      | 6±0.0 <sup>b</sup> | 8±0.0 <sup>a</sup> | 8±0.0 <sup>a</sup>  | 8±0.0 <sup>a</sup> | 10±0.0 <sup>a</sup> |
| <i>L. monocytogenes</i> | 8±0.0 <sup>a</sup>   | 10±0.0 <sup>a</sup> | >10 <sup>a</sup>        | >10 <sup>a</sup>   | >10 <sup>a</sup>   | >10 <sup>a</sup>    | >10 <sup>a</sup>   | >10 <sup>a</sup>    |
| <i>S. typhimurium</i>   | 10±0.0 <sup>a</sup>  | 10±0.0 <sup>a</sup> | >10 <sup>a</sup>        | >10 <sup>a</sup>   | >10 <sup>a</sup>   | >10 <sup>a</sup>    | >10 <sup>a</sup>   | >10 <sup>a</sup>    |
| <i>E. coli</i> O157:H7  | 10±0.0 <sup>a</sup>  | 10±0.0 <sup>a</sup> | >10 <sup>a</sup>        | >10 <sup>a</sup>   | >10 <sup>a</sup>   | >10 <sup>a</sup>    | >10 <sup>a</sup>   | >10 <sup>a</sup>    |

\* The following concentrations were used in the present study: 0.5, 1, 2, 4, 6, 8 and 10 mg/ml

\* Columns representing different values are labeled with different letters ( $p < 0.05$ )

**Table 2:** Antibacterial effect of methanolic extracts of *Falcaria vulgaris*, *Allium rotundum*, *Tragopogon graminifolius*, and *Mentha longifolia* using agar disk diffusion assay

| Bacteria                | Inhibition zone (mm)    |                         |                        |                        |
|-------------------------|-------------------------|-------------------------|------------------------|------------------------|
|                         | <i>M. longifolia</i>    | <i>T. graminifolius</i> | <i>A. rotundum</i>     | <i>F. vulgaris</i>     |
| <i>S. aureus</i>        | 8.88±0.01 <sup>b</sup>  | 4.26±0.05 <sup>b</sup>  | 3.14±0.00 <sup>b</sup> | 3.14±0.00 <sup>b</sup> |
| <i>B. subtilis</i>      | 12.31±0.01 <sup>a</sup> | 9.36±0.06 <sup>a</sup>  | 6.11±0.06 <sup>a</sup> | 3.14±0.00 <sup>b</sup> |
| <i>B. cereus</i>        | 12.45±0.03 <sup>a</sup> | 10.56±0.01 <sup>a</sup> | 7.04±0.01 <sup>a</sup> | 4.24±0.01 <sup>a</sup> |
| <i>L. monocytogenes</i> | 7.05±0.06 <sup>b</sup>  | 3.14±0.07 <sup>b</sup>  | 3.14±0.05 <sup>b</sup> | ND                     |
| <i>S. typhimurium</i>   | 3.14±0.02 <sup>c</sup>  | 3.14±0.01 <sup>b</sup>  | 3.14±0.02 <sup>b</sup> | ND                     |
| <i>E. coli</i> O157:H7  | 3.14±0.01 <sup>c</sup>  | 3.14±0.05 <sup>b</sup>  | 3.14±0.03 <sup>b</sup> | ND                     |

\* ND: Not detected (the inhibition zone was not observed)

\* Columns representing different values are labeled with different letters ( $p < 0.05$ )

Hamedan province of Iran and examined its antibacterial effect by disk diffusion method. These authors found that methanolic extract of *F. vulgaris* (concentration of 10 mg/ml) had inhibition zone of 7-10 mm against various food-borne pathogens such as *B. cereus*, *E. coli*, *S. aureus*, *Pseudomonas aeruginosa*, *Proteus vulgaris*, and *Bacillus megateriu*. Moreover, Rasooli and Rezaei (2002) reported that *M. longifolia* essential oil had inhibition zone of 8-12 mm against *S. aureus*, *B. subtilis*, *B. cereus*, *L. monocytogenes*, *S. typhimurium*, and *E. coli* O157:H7, which is in agreement with our findings. Also, these researchers showed high antibacterial activities of these extracts on Gram-positive and Gram-negative bacteria, which are in agreement with the results of the present study. However, reported different antibacterial activities of plant extracts could be due to the different bacterial strains, media, and chemical composition of the extracts (Hyldgaard et al., 2012; Lv et al., 2011).

The antioxidant activity of plant leave extracts is mostly attributed to the active compounds of phenolic fraction present in them (Akrami et al., 2015; Wannas et al., 2010). In the present research, all studied plants were obtained at the same time during the flowering stage in order to prevent climatic, edaphic as well as developmental influence on the antioxidant activity. As previously reported, antioxidant activity of *M. longifolia* extract could be related to its phenolic content like phenolic acid,

rosmarinic acid, and polyphenols (Asekun et al., 2007; Gulluce et al., 2007; Mkaddem et al., 2009). Moreover, the remarkable difference in the antioxidant property of the investigated plant extracts is probably due to variability in the nature and concentration of main groups of chemical compositions (Lv et al., 2011). Similar with our findings, Gulluce et al. (2007) found that IC<sub>50</sub> of methanolic extract of *M. longifolia* was 0.81 mg/ml. In another study by Mimica-Dukic et al. (1999), the antioxidant activity of *M. longifolia* extract was reported 0.85 mg/ml. Assadpour et al. (2016) found that the antioxidant activity of *A. rotundum* extract was 0.24 mg/ml, which is good in agreement with our findings. On the other hand, the remarkable antioxidant activity of methanolic *T. graminifolius* extract which encompasses a key role on various medicinal properties of this plant, including protective and healing function on peptic and duodenal ulcer and also wound healing and skin repairing activity (Farzaei et al., 2014a, b). Indeed, phenolic compounds are one of the main constituents of *Tragopogon* species, which has critical role in their antioxidant activities. Farzaei et al. (2014b) showed high content of total phenol (0.560±0.85 mg/g gallic acid equivalents) in ethanolic extract from *T. graminifolius* aerial part. The potential antioxidant activity of the methanolic extracts of *F. vulgaris* showed a high effective free radical scavenging in the DPPH assay, which is good in agreement



with previous studies (Jaberian et al., 2013; Monfared et al., 2012; Shafaghat, 2010). Jaberian et al. (2013) collected *F. vulgaris* from Hamedan province in the West of Iran and examined its antioxidant property by DPPH assay. Based on their findings, the IC<sub>50</sub> value of the methanolic extract of *F. vulgaris* was 0.1273 mg/ml. Monfared et al. (2012) reported that IC<sub>50</sub> in DPPH, reducing power, and total antioxidant capacity of methanolic extract of *F. vulgaris* were 97.36±0.29, 200.118, and 0.147 µg/ml, respectively.

## Conclusion

Based on the results of the present study, *F. vulgaris*, *A. rotundum*, *T. graminifolius*, and also *M. longifolia* had antimicrobial as well as antioxidant activities that recommended to be used as preservatives in food products. However, methanolic extract of *M. longifolia* had the best antibacterial and antioxidant properties *in vitro*. Further researches are required to identify the detail phytochemical compounds of these plant extracts responsible for their antibacterial and antioxidant characteristic.

## Conflicts of interest

There is no conflict of interest in this study.

## Acknowledgments

This research was ethically approved by the local institutional review board. The researcher is thankful to the taxonomist Dr. Seyed Mohammad Masoumi (Faculty of Agriculture, Razi University, Kermanshah, Iran) for his great assistance in the identification of the plant used in the study. This study was self-funded.

## References

- Akrami F., Rodríguez-Lafuente A., Bentayeb K., Pezo D., Ghalebi S., Nerín C. (2015). Antioxidant and antimicrobial active paper based on *Zataria* (*Zataria multiflora*) and two cumin cultivars (*Cuminum cyminum*). *LWT-Food Science and Technology*. 60: 929-933.
- Alves-Silva J.M., dos Santos S.M.D., Pintado M.E., Pérez-Álvarez J.A., Fernández-López J., Viuda-Martos M. (2013). Chemical composition and *in vitro* antimicrobial, antifungal and antioxidant properties of essential oils obtained from some herbs widely used in Portugal. *Food Control*. 32: 371-378.
- Alzoreky N., Nakahara K. (2003). Antibacterial activity of extracts from some edible plants commonly consumed in Asia. *International Journal of Food Microbiology*. 80: 223-230.
- Asekun O.T., Grierson D.S., Afolayan A.J. (2007). Effects of drying methods on the quality and quantity of the essential oil of *Mentha longifolia* L. subsp. *Capensis*. *Food Chemistry*. 101: 995-998.
- Assadpour S., Nabavi S.M., Nabavi S.F., Dehpour A.A., Ebrahimpour M.A. (2016). *In vitro* antioxidant and antihemolytic effects of the essential oil and methanolic extract of *Allium rotundum* L. *European Review for Medical and Pharmacological Sciences*. 20: 5210-5215.
- Benkeblia N. (2004). Antimicrobial activity of essential oil extracts of various onions (*Allium cepa*) and garlic (*Allium sativum*). *LWT-Food Science and Technology*. 37: 263-268.
- Carović-Stanko K., Orlić S., Politeo O., Strikić F., Kolak I., Milos M., Satovic Z. (2010). Composition and antibacterial activities of essential oils of seven *Ocimum taxa*. *Food Chemistry*. 119: 196-201.
- Conforti F., Menichini F., Formisano C., Rigano D., Senatore F., Arnold N.A., Piozzi F. (2009). Comparative chemical composition, free radical-scavenging and cytotoxic properties of essential oils of six *Stachys* species from different regions of the Mediterranean area. *Food Chemistry*. 116: 898-905.
- Farzaei M.H., Khanavi M., Moghaddam G., Dolatshahi F., Rahimi R., Shams-Ardekani M.R., Amin G., Hajimahmoodi M. (2014a). Standardization of *Tragopogon graminifolius* DC. extract based on phenolic compounds and antioxidant activity. *Journal of Chemistry*. 2014: 1-7.
- Farzaei M.H., Rahimi R., Attar F., Siavoshi F., Saniee P., Hajimahmoodi M., Mirnezami T., Khanavi M. (2014b). Chemical composition, antioxidant and antimicrobial activity of essential oil and extracts of *Tragopogon graminifolius*, a medicinal herb from Iran. *Natural Product Communications*. 9: 121-124.
- Ghasemi Pirbalouti A., Izadi A., Malek Poor F., Hamed B. (2016). Chemical composition, antioxidant and antibacterial activities of essential oils from *Ferulago angulata*. *Pharmaceutical Biology*. 54: 2515-2520.
- Gulluce M., Sahin F., Sokmen M., Ozer H., Daferera D., Sokmen A., Polissiou M., Adiguzel A., Ozkan H. (2007). Antimicrobial and antioxidant properties of the essential oils and methanol extract from *Mentha longifolia* L. ssp. *longifolia*. *Food Chemistry*. 103: 1449-1456.
- Gyawali R., Ibrahim S.A. (2014). Natural products as antimicrobial agents. *Food Control*. 46: 412-429.
- Hussain A.I., Anwar F., Sherazi S.T.H., Przybylski R. (2008). Chemical composition, antioxidant and antimicrobial activities of basil (*Ocimum basilicum*) essential oils depends on seasonal variations. *Food Chemistry*. 108: 986-995.
- Hyldgaard M., Mygind T., Meyer R.L. (2012). Essential oils in food preservation: mode of action, synergies, and interactions with food matrix components. *Frontiers in Microbiology*. 3: 1-24.
- Jaberian H., Piri K., Nazari J. (2013). Phytochemical composition and *in vitro* antimicrobial and antioxidant activities of some medicinal plants. *Food Chemistry*. 136: 237-244.
- Jay J.M., Loessner M.J., Golden D.A. (2005). Modern food microbiology. 7<sup>th</sup> edition. Springer Science, New York.
- Lv F., Liang H., Yuan Q., Li C. (2011). *In vitro* antimicrobial effects and mechanism of action of selected plant essential oil combinations against four food-related microorganisms. *Food Research International*. 44: 3057-3064.
- Mimica-Dukic N., Popovic M., Jakovljevic V., Szabo A., Gašić O. (1999). Pharmacological studies of *Mentha longifolia* phenolic extracts. II. Hepatoprotective activity. *Pharmaceutical Biology*. 37: 221-224.
- Mkaddem M., Bouajila J., Ennajar M., Lebrihi A., Mathieu F., Romdhane M. (2009). Chemical composition and antimicrobial and antioxidant activities of *Mentha* (*longifolia* L. and *viridis*) essential oils. *Journal of Food Science*. 74: 358-363.
- Monfared K.E., Rafiee Z., Jafari S. (2012). Phenolic content and antioxidant activity of *Falcaria vulgaris* extracts. *Analytical Chemistry Letters*. 2: 159-170.
- Motamed S.M., Naghibi F. (2010). Antioxidant activity of some edible plants of the Turkmen Sahra region in Northern Iran. *Food Chemistry*. 119: 1637-1642.
- Pal D., Banerjee S., Ghosh A.K. (2012). Dietary-induced cancer prevention: an expanding research arena of emerging diet related to healthcare system. *Journal of Advanced Pharmaceutical Technology and Research*. 3: 16-18.

- Park M., Bae J., Lee D.S. (2008). Antibacterial activity of [10]-gingerol and [12]-gingerol isolated from ginger rhizome against periodontal bacteria. *Phytotherapy Research*. 22: 1446-1449.
- Rasooli I., Rezaei M.B. (2002). Bioactivity and chemical properties of essential oils from *Zataria multiflora* Boiss and *Mentha longifolia* (L.) Huds. *Journal of Essential Oil Research*. 14: 141-146.
- Runyoro D., Ngassapa O., Vagionas K., Aligiannis N., Graikou K., Chinou I. (2010). Chemical composition and antimicrobial activity of the essential oils of four *Ocimum* species growing in Tanzania. *Food Chemistry*. 119: 311-316.
- Sakunpak A., Panichayupakaranant P. (2012). Antibacterial activity of Thai edible plants against gastrointestinal pathogenic bacteria and isolation of a new broad spectrum antibacterial polyisoprenylated benzophenone, chamuangone. *Food Chemistry*. 130: 826-831.
- Schinella G., Tournier H., Prieto J., De Buschiazzo P.M., Rios J. (2002). Antioxidant activity of anti-inflammatory plant extracts. *Life Sciences*. 70: 1023-1033.
- Shafaghat A. (2010). Free radical scavenging and antibacterial activities, and GC/MS analysis of essential oils from different parts of *Falcaria vulgaris* from two regions. *Natural Product Communications*. 5: 981-984.
- Shahbazi Y. (2015). Antilisterial effects of *Ziziphora clinopodioides* essential oil and nisin in milk. *Journal of Pure and Applied Microbiology*. 9: 1993-1999.
- Shahbazi Y., Shavisi N. (2016). Interactions of *Ziziphora clinopodioides* and *Mentha spicata* essential oils with chitosan and ciprofloxacin against common food-related pathogens. *LWT-Food Science and Technology*. 71: 364-369.
- Shahbazi Y., Shavisi N., Karami N., Kakaei S. (2015). Chemical composition and *in vitro* antibacterial activity of *Ferulago angulata* (Schlecht.) Boiss essential oil. *Pharmaceutical Sciences*. 21: 6-11.
- Shahbazi Y., Shavisi N., Mohebi E. (2016). Potential application of *Ziziphora clinopodioides* essential oil and nisin as natural preservatives against *Bacillus cereus* and *Escherichia coli* O157:H7 in commercial barley soup. *Journal of Food Safety*. 36: 435-441.
- Wannes W.A., Mhamdi B., Sriti J., Jemia M.B., Ouchikh O., Hamdaoui G., Kchouk M.E., Marzouk B. (2010). Antioxidant activities of the essential oils and methanol extracts from myrtle (*Myrtus communis* var. *italica* L.) leaf, stem and flower. *Food and Chemical Toxicology*. 48: 1362-1370.
- Yang X.J., Zhang M., Zhu H.M., Tang Z., Zhao D.D., Li B.Y., Gabriel A. (2016). Epidemiological study: correlation between diet habits and constipation among elderly in Beijing region. *World Journal of Gastroenterology*. 22: 8806-8809.