



Parasitic Contamination in Five Leafy Vegetables Collected from Open Marketplaces in Giza, Egypt

N.M. El-Sayed ^{1*}, S.S. Gawdat ¹, H.S. El-Kholy ¹, A. Elmosalamy ²

1. Medical Parasitology Department, Research Institute of Ophthalmology, Giza, Egypt

2. Medical Student, Faculty of Medicine, Cairo University, Cairo, Egypt

HIGHLIGHTS

- Leafy vegetables had a very high parasite contamination rate (86%) in Giza, Egypt.
- Rate of contamination by protozoan parasites was higher than rate of contamination by helminth parasites.
- *Entamoeba histolytica/dispar* was the most encountered parasite with an estimated prevalence reaching 40.6%.

Article type

Original article

Keywords

Vegetables
Parasites
Helminths
Parasitology
Public Health
Egypt

Article history

Received: 6 May 2022

Revised: 7 Oct 2022

Accepted: 13 Nov 2022

ABSTRACT

Background: Leafy vegetables could be contaminated with parasitic infective stages, making them possible sources of human parasitic infections. This study aimed to assess parasites contamination level among five different leafy vegetables collected from open marketplaces in Giza, Egypt.

Methods: Hundred vegetable samples were collected from dill, parsley, coriander, rocca, and mint (20 of each). Samples were processed using sedimentation and concentration methods and the sediments were examined as wet smears either unstained or stained with Lugol's iodine. Also, modified trichrome and Ziehl-Neelsen stains were used to detect *Microsporidia* spores and coccidian oocysts. Statistical software SPSS version 20 was used for data analysis.

Results: A total of 86% of the examined samples were found contaminated with parasite's eggs, cysts, oocysts, and larvae. Coriander was the most contaminated vegetable with contamination rate of 95% while dill was the least contaminated (80%). Also, protozoan parasites contamination rate (77.1%) was higher than helminths parasites contamination rate (22.9%). *Entamoeba histolytica/dispar* was the most prevalent parasite (40.6%) followed by *Blastocystis hominis* (39.5%), *Ascaris lumbricoides* (18.6%), *Cyclospora* spp. (15.11%), *Giardia lamblia* (11.6%), *E. coli* (9.3%); while *Cryptosporidium* spp., *Iodamoeba bütschlii*, *Trichuris trichiura*, *Enterobius vermicularis*, *Fasciola*, and hookworms' eggs were the least prevalent parasites (1.1%).

Conclusion: Leafy vegetables in Giza, Egypt had an extremely high parasite contamination rate. So, consuming such vegetables without proper washing facilitates transmission of parasitic infections which could pose a major health risk to Egyptians.

© 2023, Shahid Sadoughi University of Medical Sciences. This is an open access article under the Creative Commons Attribution 4.0 International License.

Introduction

Many parasites may be transmitted through food. The most prevalent food-borne parasites in underdeveloped

countries are intestinal parasites. Intestinal parasite infections pose a serious threat to public health all over

* Corresponding author (N.M. El-Sayed)

✉ E-mail: nag.elsaka@yahoo.com

ORCID ID: <https://orcid.org/0000-0001-6880-2147>

To cite: El-Sayed N.M., Gawdat S.S., El-Kholy H.S., Elmosalamy A. (2023). Parasitic contamination in five leafy vegetables collected from open marketplaces in Giza, Egypt. *Journal of Food Quality and Hazards Control*. 10: 13-20.

the world, particularly in children where they result in diarrhea, malnutrition, anemia as well as some developmental disorders (El-Sayed and Ramadan, 2017a).

Leafy vegetables can help to improve nutritional status and avoid a variety of ailments. They are a rich source of vitamins, minerals, and dietary fibers, making them an essential component of a balanced human diet (Traoré et al., 2021). However, various human populations' eating habits and hygiene standards influence the chances of acquiring a parasite infection through food. Eating unwashed raw vegetables has been associated with the acquisition of several intestinal parasites, such as protozoa cysts and oocysts or helminths' eggs and larvae, which in turn increase the risk of food-borne illnesses (El-Sayed et al., 2021).

Parasitic contamination of vegetables has been related to several factors such as the use of untreated wastewater for irrigation, soil contamination with humans and animals' stools, and the use of improperly managed animals manure as fertilizer (Bekele et al., 2017). A moist environment is necessary for the growth of vegetables, and this moisture also aids in the development of several parasite species (Abougrain et al., 2010). Vegetables may become contaminated while being harvested, transported, stored, marketed, and otherwise processed in an unhygienic manner (El-Sayed et al., 2021).

Vegetables contamination by different parasites has been documented in several studies from different countries. The prevalence of contamination varies widely among different regions ranging from 15.1% in the United Arab Emirates (El Bakri et al., 2020) to 100% in Yemen (Al-Sanabani et al., 2016). The variations may depend on geographical locations, conditions of climate and environments, the type of vegetables examined, the used sample size and the used techniques for identification of the parasites in the samples (Tefera et al., 2014).

There have been relatively few studies focused on evaluating the parasite contamination of vegetables in various Egyptian regions (Ahmed et al., 2020; Eteawa et al., 2017; Hassan et al., 2012). Determining the level of vegetables contamination may aid in the development of effective anti-parasitic control strategies to promote health and offer protection from diseases. In this regard, the present investigation was planned to assess parasites contamination level among five different leafy vegetables collected from open marketplaces in Giza, Egypt.

Materials and methods

Study area

This cross-sectional study was carried out between July and October 2020 using randomly selected vegetable samples from open marketplaces in Giza city, Egypt. Giza, a city of the Greater Cairo metropolis, is situated on the West bank of the Nile, 4.9 km Southwest of Central Cairo. It has a total area of 1,580 km². It has a population of 2.4 million. It has many areas which are under small and large-scale vegetable cultivation. The average maximum temperature in this agricultural region during the summer is 35.5 °C, while the average low is around 23.3 °C with an average humidity of 48%.

Collection of vegetable samples

In this study, 100 samples from five different green leafy vegetables-20 samples of each-were examined. The studied vegetables were dill (*Anethum graveolens* L.), parsley (*Petroselinum crispum*), coriander (*Coriandrum sativum*), rocca (*Eruca sativa*), and mint (*Mentha piperita* L.). These vegetables were chosen because they are regularly consumed raw and are widely available and sold in every market of the study area. Vegetables were purchased from randomly selecting street sellers and vegetable retailers. Following that, each sample was placed in a distinct plastic bag and identified by a unique number. Within 24 h of sample collection, each sample was processed and examined for parasites detection.

Samples processing

To separate the parasites from the vegetables, 100 g of each vegetable were soaked in 1,000 ml of physiological saline solution (0.9% sodium chloride) with agitation for 15 min. The washing solution was then let to stand on the bench overnight for optimal sedimentation. The supernatant was decanted on the next day, and the residual washing water was filtered through a strainer with a pore size of 300 µm to eliminate unwanted debris before being allowed to sediment for another hour. Then, the supernatant was discarded once more. The remaining sediment was collected into conical tubes and centrifuged at 3,000×g for 15 min to concentrate the parasitic stages. The supernatant was carefully collected after centrifugation, and the sediments were examined for parasite ova/cysts detection.

Detection of parasites

Four slides were prepared from each sample to increase

the chance of detecting parasites. The first slide was inspected as unstained wet smear, whereas the second one was stained with Lugol's Iodine. Both slides were examined for parasitic protozoa and helminths using 10× and 40× objectives of a light microscope (Zeiss, Axio Scope.A1, Jena, Germany). On other slides, a thin smear of the sediment was applied, air dried, and then fixed for 3 min with methanol. The slides were stained with trichrome stain for detection of *Microsporidium* spp. spores, and modified Ziehl-Neelsen for detection of coccidian oocysts as *Cryptosporidium* spp., and *Cyclospora* spp. (WHO, 2019). Slides were then examined microscopically by oil immersion lens (100×). The microscopic examination was done 3 times on each slide by independent investigators for confirmation. Parasitic stages were recognized depending on their morphological characteristics according to WHO (2019).

Statistical analysis

Statistical software SPSS version 20 was used for data analysis. Calculation of the contamination rate percentage was done by dividing the number of positive samples on the number of total vegetables and the resulting number was multiplied by 100. Also, mean density (parasites/100 g of vegetables) was calculated. Chi-square test was used to compare the rate of parasite contamination in the studied vegetables, compare the vegetables contamination by a single parasite versus multiple parasites and also, compare the vegetables contamination by protozoa versus helminths. The p values <0.05 were considered as significant.

Results

Totally, 86 out of 100 studied vegetable samples were contaminated with different parasitic eggs, cysts, oocysts,

and larvae (Figures 1 and 2). Coriander was the most contaminated vegetable with contamination rate of 95% (19 out of 20), while dill was the least contaminated (16 out of 20; 80%). Parsley, rocca, and mint gave the same contamination rates of 85% (17 out of 20). There was no significance difference between the rate of parasite contamination among five types of vegetables ($\chi^2=1.9934$; $p=0.736981$).

Out of the overall contaminated samples, 36.1% had one type of parasite and the remaining 63.9% had multiple parasitic stages: double (41.86%), triple (16.27%), quadruple (4.65%), or quintuple (1.16%) parasites. There was no statistically significant difference even though the rate of vegetables contamination by multiple parasites was higher than rate of contamination by single parasite ($\chi^2=9.3527$; $p=0.052864$; Table 1). Detected parasites included 128 (77.1%) protozoan cysts/oocysts, and 38 (22.9%) helminthic eggs, larvae, and worms. Although, the rate of contamination by protozoan parasites was higher than rate of contamination by helminth parasites, there was no significance difference ($\chi^2=5.3846$; $p=0.250063$; Table 2).

Regarding the parasite prevalence among the studied vegetables, *E. histolytica/dispar* was the most encountered parasite with an estimated prevalence reaching 40.6% followed by *B. hominis* (39.5%), *A. lumbricoides* (18.6%), *Cyclospora* spp. (15.11%), *G. lamblia* (11.6%), and *E. coli* (9.3%). While *Cryptosporidium* spp., *I. bütschlii*, *T. trichiura*, *E. vermicularis*, *Fasciola*, and hookworms' eggs were the least prevalent (1.1%) parasites in the studied vegetables. Other detected parasites included *T. gondii*, *H. nana*, *S. stercoralis*, *E. nana*, *B. coli*, *C. mesinili*, *Microsporidia* spp., *H. diminuta*, and *Toxocara* spp. with prevalence rates ranging from 8.1 to 2.3% (Table 3).

Table 1: Intensity of parasitic contamination in studied vegetables samples from Giza, Egypt

Studied Vegetables	No. of contaminated samples	Single Contamination No. (%)	No. of mixed contamination				Total No. (%)
			Double cont.	Triple cont.	Quadruple cont.	Quintuple cont.	
Dill	16	6 (37.5)	6	3	1	0	10 (62.5)
Parsley	17	11 (64.7)	3	3	0	0	6 (35.3)
Coriander	19	5 (26.3)	13	1	0	0	14 (73.7)
Rocca	17	6 (35.3)	6	3	2	0	11 (64.7)
Mint	17	3 (17.6)	8	4	1	1	14 (82.4)
Total	86	31 (36.1)	36 (41.86%)	14 (16.27%)	4 (4.65%)	1 (1.16%)	55 (63.9)

$\chi^2=9.3527$; $p=0.052864$

Table 2: Prevalence of protozoan and helminths contamination in studied vegetables samples from Giza, Egypt

Studied Vegetables	Total No. of parasites	Protozoa Contamination No. (%)	Helminths Contamination No. (%)
Dill	31	26 (83.8)	5 (16.1)
Parsley	26	23 (88.5)	3 (11.5)
Coriander	34	26 (76.5)	8 (23.5)
Rocca	35	23 (65.7)	12 (34.3)
Mint	40	30 (75)	10 (25)
Total	166	128 (77.1)	38 (22.9)

$\chi^2=5.3846$; $p=0.250063$

Table 3: Distribution and prevalence of the detected parasites in contaminated vegetables samples from Giza, Egypt

Detected parasites	No. of positive vegetables samples (86)					Total No.	Prevalence %	Mean density
	Dill (16)	Parsley (17)	Coriander (19)	Rocca (17)	Mint (17)			
Protozoa								
<i>Entamoeba histolytica/dispar</i> cyst	6	8	6	5	10	35	40.6	7
<i>Entamoeba coli</i> cyst	1	2	2	3	0	8	9.3	1.6
<i>Endolimax nana</i> cyst	0	0	1	2	0	3	3.4	0.6
<i>Iodamoeba bütschlii</i> cyst	1	0	0	0	0	1	1.1	0.2
<i>Chilomastix mesinili</i> cyst	0	0	1	0	1	2	2.3	0.4
<i>Balantidium coli</i> cyst	0	0	0	2	1	3	3.4	0.6
<i>Giardia. lamblia</i> cyst	2	4	1	-	3	10	11.6	2
<i>Cyclospora</i> spp. oocyst	2	1	2	4	4	13	15.11	2.6
<i>Cryptosporidium</i> oocyst	0	0	0	0	1	1	1.1	0.2
<i>Microsporidia</i> spores	2	0	0	0	0	2	2.3	0.4
<i>Blastocystis hominis</i> cyst	9	6	9	7	3	34	39.5	6.8
<i>Toxoplasma gondii</i> oocyst	0	2	2	0	3	7	8.1	1.4
Unsporulated oocyst	3	0	2	0	4	9	10.4	1.8
Total No.	26	23	26	23	30	128		
Helminths								
<i>Ascaris lumbricoides</i> egg	1	1	3	8	3	16	18.6	3.2
<i>Trichuris trichiura</i> egg	0	0	0	0	1	1	1.1	0.2
<i>Hymenolepis nana</i> egg	1	1	0	1	3	6	6.9	1.2
<i>Hymenolepis diminuta</i> egg	1	0	1	0	0	2	2.3	0.4
<i>Enterobius vermicularis</i> egg	1	0	0	1	0	2	2.3	0.4
<i>E. vermicularis</i> worm	0	0	1	0	0	1	1.1	0.2
<i>Strongyloides stercoralis</i> larvae	0	1	2	2	1	6	6.9	1.2
<i>Toxocara</i> spp. egg	1	0	1	0	0	2	2.3	0.4
<i>Fasciola</i> egg	0	0	0	0	1	1	1.1	0.2
Hookworm egg	0	0	0	0	1	1	1.1	0.2
Total No.	5	3	8	12	10	38	9.6	3.2
Total No. of detected parasites						166		

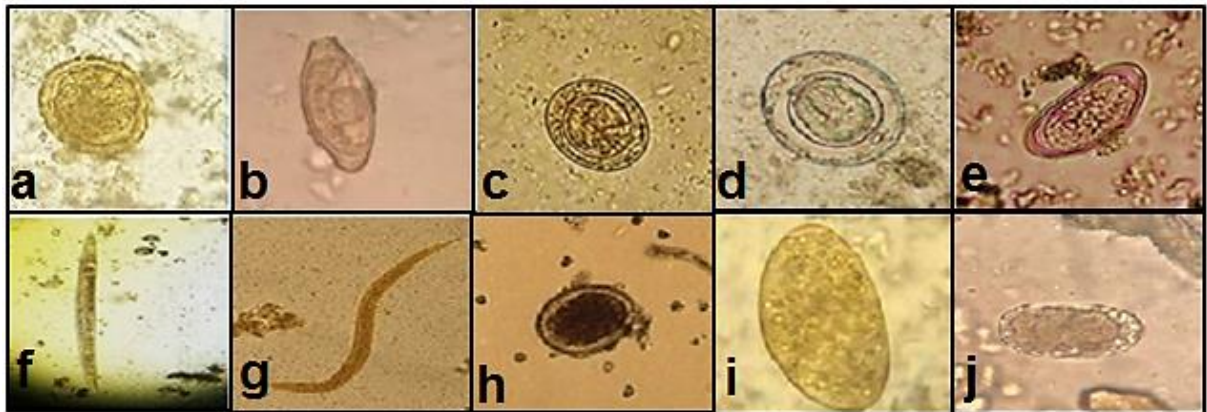


Figure 1: Detected helminthic eggs, larvae, and worms in the studied vegetables samples from Giza, Egypt: a: *Ascaris lumbricoides* egg; b: *Trichuris trichiura* egg; c: *Hymenolepis nana* egg; d: *Hymenolepis diminuta* egg; e: *Enterobius vermicularis* egg; f: *E. vermicularis* worm; g: *Strongyloides stercoralis* larva; h: *Toxocara* spp. egg; i: *Fasciola* egg; j: Hookworm egg

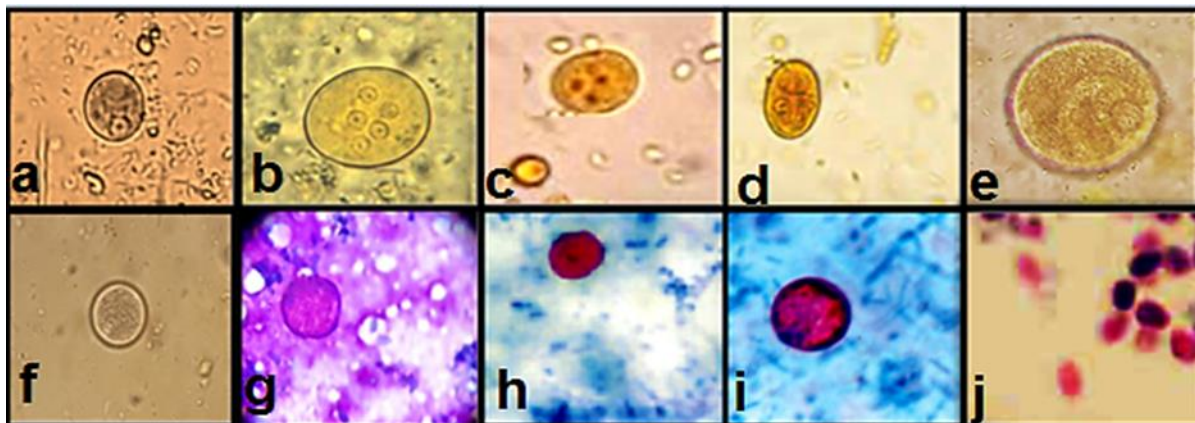


Figure 2: Detected protozoan cysts/oocysts in the studied vegetables samples from Giza, Egypt: a: *Entamoeba histolytica/dispar* cyst; b: *Entamoeba coli* cyst; c: *Endolimax nana* cyst; d: *Giardia lamblia* cyst; e: *Balantidium coli* cyst; f: *Toxoplasma gondii* oocyst; g: *Blastocystis hominis* cyst; h: *Cyclospora* spp. oocyst; i: *Cryptosporidium* spp. oocyst; j: *Microsporidia* spores

Discussion

Most Egyptians have a dietary practice of eating raw leafy vegetables such as dill, parsley, coriander, rocca, and mint with their meals to provide the body with a variety of nutrients and vitamins. The easier attachment of infective parasitic stages to the leaves of raw vegetables always raises the danger of vegetables-borne parasitic diseases (El-Sayed et al., 2021). Therefore, the current study was done to assess the level of contamination and the prevalence of various intestinal parasites in the most prevalent leafy vegetables consumed by Egyptians in Giza city.

The current study revealed a high level of contamination (86%) in the examined vegetables. This contamination could be due to the leaves' ability to retain parasites between them, as well as their irregular surfaces, which facilitate easier parasite attachment. Also, vegetable samples were gathered in the summer months when parasite dissemination and prevalence are often high, which contributed to the study's high contamination rate. Parasitic contamination of vegetables was documented in other Egyptian studies that were conducted in various cities. Amongst Egyptian cities, Assiut city had the highest contamination rate at 84.1% (Ahmed et al., 2020), while Alexandria city had the lowest contamination rate at 19.4% (Hassan et al., 2012). Moreover, our findings agreed with those of several researches conducted in various countries where the parasite contamination rate of vegetables ranged from a lower contamination rate of 15.1% (El Bakri et al., 2020) to a higher contamination rate of 100% (Al-Sanabani et al., 2016). This might be due to variations in the climatic

conditions, investigated vegetables, and research methodologies between this study and other studies.

In this study, rate of contamination by protozoan parasites (77.1%) was higher than rate of contamination by helminth parasites (22.9%). This is in agreement with the findings of Elmajdoub et al. (2017), who found that the contamination rate was 79.4% for protozoan cysts and 47.6% for helminth parasites. The presence of multiple species of medically important parasites in the studied vegetables certainly reflects the high level of contamination and the persistence of intestinal parasitic infection in Giza city. This could result in widespread intestinal parasite infections among humans (Youssef and Uga, 2014). In the present study, *E. histolytica/dispar* has shown the highest percentage (40.6%) as it was detected in all studied vegetables. Such a high percentage was observed in a study from Nigeria in which *E. histolytica* was the only detected protozoan parasite (Oladele et al., 2022), however, the percentage was lower in Saudi Arabia report (3.1%; Al-Binali et al., 2006). Variations in geographic distribution and the cysts' ability to survive in cold, moist circumstances for a long time may be responsible for the predominance of *E. histolytica/dispar*.

B. hominis was the second most prevalent contaminant in the studied vegetables which recorded 39.5%. The obtained result agreed with the results of Al Nahhas and Aboualchamat (2020) in Syria (29.5%). However, Etewa et al. (2017) recorded a lower rate (3.8%) in Sharkyia, Egypt. The high prevalence of *B. hominis* might be attributed to the use of trichrome stain as a diagnostic tool in the present study in contrast to the other studies. Other parasites detected in this study included

Cyclospora spp. (15.11%), *Microsporidia* (2.3%), and *Cryptosporidium* (1.1%) which are leading cause of diarrhea worldwide with significant morbidity and mortality particularly in the immune-compromised individuals. However, much higher incidence rate of these parasites reported in another Egyptian study by Said (2012) who recorded *Cryptosporidium* oocysts in 29.3% of raw vegetables followed by *Microsporidia* (25.3%) and *Cyclospora* (21.3%). The difference between this study and others might be due to differences in the studied vegetables, sample size, and used methods.

Result of this research, showing 11.6% of vegetables were *G. lamblia*-contaminated, is consistent with previous research in Egypt, done in Alexandria, by Said (2012) and Hassan et al. (2020) who reported prevalence's rates of 6.7 and 8.8%, respectively. Similar finding was drawn from Libyan research, where 10% of all examined vegetable samples had *G. lamblia* cysts (Abougrain et al., 2010). Furthermore, this result is compatible with the percentage of Egyptians infected with giardiasis, which ranges from 10 to 34.6% (Youssef and Uga, 2014). This prevalence may be attributed to the *Giardia*'s ability to survive under cool and moist conditions as well as its resistance to chlorine in drinking water. *B. coli* was also found in 3.4% of the tested vegetables. However, it was the most prevalent parasite contaminant in the studies conducted in Cameroon (25.6%; Akoachere et al., 2018) and Ghana (13.6%; Kudah et al., 2018). *B. coli*-contaminated vegetables may be caused by contaminating irrigation water with humans and pigs feces (Kudah et al., 2018). Other detected intestinal protozoa in this study were *E. coli* (9.3%), *E. nana* (3.4%), *I. bütschlii* (1.1%), and *C. mesinili* (2.3%). The presence of these non-pathogenic parasites in vegetable samples indicated a low level of sanitation, as the vegetables were contaminated with human feces.

We found that 8.1% of vegetables were contaminated with *T. gondii* oocysts due to substantial contamination of the Egyptian environment by stray cats feces. This result agrees with that found by Marček et al. (2018) where 6.7% of examined lettuce was contaminated with *T. gondii* oocysts. However, other studies relied on the detection of *Toxoplasma* DNA on the examined vegetables which certainly will give much higher contamination rates (Marques et al., 2020). Toxoplasmosis is considered a significant health problem among Egyptian population with high sero-prevalence levels ranging from 33.67% (El-Sayed et al., 2016a) to 54% (El-Sayed et al., 2016b).

Vegetables that are frequently fertilized with natural fertilizers such as animal excreta are more liable to contamination. *A. lumbricoides*, *Toxocara* spp., *T. trichiura*, and *Ancylostoma duodenale* are among the parasites whose eggs are present in this manure. Over

time, these eggs will mature and eventually reach infective stages, which might lead to helminth infections. Regarding helminthic infections detected in this study, the parasite with the highest prevalence was *A. lumbricoides* (18.6%). This might be because the female parasite producing a large number of eggs that are resistant to harsh environments (Roberts, 2009) and also, the embryonated egg's albuminoid coat characteristic that allows the parasite to adhere easily to vegetables. The prevalence of *A. lumbricoides* is consistent with findings from earlier studies in Egypt (20.3%; Said, 2012), Ethiopia (20.8%; Bekele et al., 2017), and Pakistan (26.7%; Khan et al., 2017). Although, prevalence of hookworm and *T. trichiura* seems to be low (1.1%) in this research, it could indicate fecal pollution of the environment and inadequate sewage disposal system. Detection of *Toxocara* eggs in the studied vegetables with a rate of 2.3% might be related to the substantial contamination of the Egyptian environment by stray dogs and cats feces as well as the eggs' resistance to extreme environmental conditions, which may survive for up to ten years. The prevalence rate of *Toxocara* eggs in this study (2.3%) was consistent with what reported by Punsawad et al. (2019) in Thailand (2.6%). Many researches have documented the high prevalence of human toxocariasis, either visceral (El-Sayed and Ramadan, 2017b) or ocular (El-Sayed and Masoud, 2019) in Egypt.

H. nana was the second most frequent helminthic contamination in the studied vegetables, accounting for 6.9%. This percentage was comparable to that recorded in Libya (7.9%) (Elmajdoub et al., 2017). On contrast, other studies showed higher rate in Ethiopia (15.56%; Bekele et al., 2017) and lower rate as detected in Qazvin, Iran, (0.5%; Shahnazi and Jafari-Sabet, 2010). Differences in the environmental conditions and also geographical location could explain the observed disparity. According to numerous researchers, hymenolypiasis infections are exceedingly common in Egyptian patients, with high detection rates (Youssef and Uga, 2014).

Strongyloides larvae were detected at a prevalence of 6.9% in this study, which could be attributable to the parasite's free-living state as well as its parasitic lifestyle. The preponderance of *Strongyloides* is consistent with findings from earlier studies (Al Nahhas and Aboualchamat, 2020; Alemu et al., 2019; Elmajdoub et al., 2017; Mohamed et al., 2016; Punsawad et al., 2019). However, *S. stercoralis* may not pose a considerable risk to consumers because human infection is primarily transmitted to barefoot individuals through larval stages penetrating their skin rather than through eating.

According to Egyptian studies, fascioliasis seems to be hyperendemic in Nile Delta villages with the mean

prevalence of 12.8%, the highest level ever recorded in Egypt (Esteban et al., 2003). However, just 1.1% of the vegetable samples in this study were contaminated with *Fasciola* eggs. Similarly, Fawzi et al. (2004) detected *Fasciola* eggs in 2.4% of the vegetable samples from a village in Alexandria governorate, Egypt. Also, other detected helminthic contaminations in the studied vegetables were *H. diminuta* and *E. vermicularis* with a same prevalence rate of 2.3%. This ensures that vegetables are fertilized with untreated human and animal wastes. In a study in Philippines, *H. diminuta* was reported to be 1% (Vizon et al., 2019) and in another study in Southwest Ethiopia, it was reported 1.4% (Tefera et al., 2014). But the results are different from those of other researchers in Iraq (4%; Saida and Nooraldeen, 2014) and southern Ethiopia (7.7%; Bekele et al., 2017). Regarding *E. vermicularis*, Etewa et al. (2017) in Sharkiya, Egypt reported 1.4% while El Bakri et al. (2020) in United Arab Emirates, reported higher rate of 6.1%. The differences between our study and prior studies may be due to differences in geographic regions, climatological or environmental variables, use of wastewater for irrigating vegetables, sample type and size, sampling processes, intestinal parasite detection methods, cross contamination during unhygienic transportation or market handling, and socioeconomic status (Said, 2012). As long as these, variables are different, there will be a disparity in the observed results.

Conclusion

The results of this study showed that leafy vegetables had a very high parasite contamination rate in Giza, Egypt. Eating of such vegetables in raw state facilitates transmission of intestinal parasitic infections that may cause a major health risk to Egyptians. To avoid risk of vegetables-borne parasitic diseases, additional research should be done to determine the degree of contamination of the soil and irrigation water used to grow these vegetables. Nevertheless, preventing contamination continues to be the most effective strategy to reduce these diseases. Both environmental and health organizations should collaborate to improve the sanitary conditions for vegetables cultivation and consumption. Consumers should be informed about the health concerns of consumption of contaminated vegetables through media programs, as well as the need to wash and disinfect vegetables before eating.

Author contributions

N.M.E.-S proposed the research idea and the study design, collected the study samples, shared in laboratory performance of the work, interpreted the results, wrote the manuscript; S.S.G. collected the study samples, assisted in performing the laboratory work, and revised the manuscript; H.S.E.-K. and A.E. assisted in performing the laboratory work. Each author examined carefully the prepared slides separately and all agreed on the positivity of samples by microscopy. All authors read and approved the final manuscript.

Conflicts of interest

None of the authors have conflicts.

Acknowledgements

This study did not receive any specific grant from funding agencies in the public, commercial, or non -for-profit sectors.

References

- Abougrain A.K., Nahaisi M.H., Madi N.S., Saied M.M., Ghenghesh K.S. (2010). Parasitological contamination in salad vegetables in Tripoli-Libya. *Food Control*. 21: 760-762. [DOI: 10.1016/j.foodcont.2009.11.005]
- Ahmed A.A., Sayed F.G., Galal L.A., Ismail T., Gaber M.M. (2020). Detection of parasites contaminating raw consumable vegetables in Assiut city, Assiut governorate, Egypt. *Journal of the Egyptian Society of Parasitology*. 50: 557-564. [DOI: 10.21608/jesp.2020.131086]
- Akoachere J.-F.T.K., Tatsinkou B.F., Nkengfack J.M. (2018). Bacterial and parasitic contaminants of salad vegetables sold in markets in Fako Division, Cameroon and evaluation of hygiene and handling practices of vendors. *BMC Research Notes*. 11: 100. [DOI: 10.1186/s13104-018-3175-2]
- Al-Binali A.M., Bello C.S., El-Shewy K., Abdulla S.E. (2006). The prevalence of parasites in commonly used leafy vegetables in South Western, Saudi Arabia. *Saudi Medical Journal*. 27: 613-616.
- Alemu G., Mama M., Misker D., Hafu D. (2019). Parasitic contamination of vegetables marketed in Arba Minch town, southern Ethiopia. *BMC Infectious Diseases*. 19: 410. [DOI: 10.1186/s12879-019-4020-5]
- Al Nahhas S., Aboualchamat G. (2020). Investigation of parasitic contamination of salad vegetables sold by street vendors in city markets in Damascus, Syria. *Food and Waterborne Parasitology*. 21: e00090. [DOI: 10.1016/j.fawpar.2020.e00090]
- Al-Sanabani A.-W., Abd Algalil F.M., Radmana B.A., Al-Manusori R.T. (2016). Prevalence of intestinal parasites in fresh leafy vegetables in some farms at Dhamar city, Yemen. *International Journal of Medicine Research*. 1: 7-13.

- Bekele F., Tefera T., Biresaw G., Yohannes T. (2017). Parasitic contamination of raw vegetables and fruits collected from selected local markets in Arba Minch town, southern Ethiopia. *Infectious Diseases of Poverty*. 6: 19. [DOI: 10.1186/s40249-016-0226-6]
- El Bakri A., Hussein N.M., Ibrahim Z.A., Hasan H., AbuOdeh R. (2020). Intestinal parasite detection in assorted vegetables in the United Arab Emirates. *Oman Medical Journal*. 35: e128. [DOI: 10.5001/omj.2020.46]
- Elmajdoub L.O., Omar M.A., Al-Aboudy M.S., AboSheba F., Elzwawi S. (2017). Prevalence of parasitic contamination of leafy green vegetables in Misurata, Libya. *Russian Journal of Parasitology*. 40: 197-204.
- El-Sayed N.M., Abdel-Wahab M.M., Kishik S.M., Alhusseini N.F. (2016a). Do we need to screen Egyptian voluntary blood donors for toxoplasmosis?. *Asian Pacific Journal of Tropical Disease*. 6: 260-264. [DOI: 10.1016/S2222-1808(15)61027-1]
- El-Sayed N.M., Kishik S.M., Fawzy R.M. (2016b). The current status of *Toxoplasma gondii* infection among Egyptian rheumatoid arthritis patients. *Asian Pacific Journal of Tropical Disease*. 6: 797-801. [DOI: 10.1016/S2222-1808(16)61133-7]
- El-Sayed N.M., Masoud N.G. (2019). Ocular toxocarasis: a neglected parasitic disease in Egypt. *Bulletin of the National Research Centre*. 43: 146. [DOI: 10.1186/s42269-019-0185-8]
- El-Sayed N.M., Ramadan M.E. (2017a). The impact of intestinal parasitic infections on the health status of children: an overview. *Journal of Pediatric Infectious Diseases*. 12: 209-213. [DOI: 10.1055/S-0037-1603576]
- El-Sayed N.M., Ramadan M.E. (2017b). Toxocarasis in children: an update on clinical manifestations, diagnosis and treatment. *Journal of Pediatric Infectious Diseases*. 12: 222-227. [DOI: 10.1055/S-0037-1603496]
- El-Sayed N.M., Ramadan M.E.E., Masoud N.G. (2021) A step forward towards food safety from parasite infective agents. In: Babalola O.O. (Editor). Food security and safety. Springer, Cham. pp: 807-832. [DOI: 10.1007/978-3-030-50672-8_40]
- Esteban J.-G., Gonzalez C., Curtale F., Muñoz-Antoli C., Valero M.A., Bargues M.D., El-Sayed M., El-Wakeel A.A.W., Abdel-Wahab Y., Montresor A., Engels D., Savioli L., et al. (2003). Hyperendemic fascioliasis associated with schistosomiasis in villages in the Nile Delta of Egypt. *American Journal of Tropical Medicine and Hygiene*. 69: 429-437.
- Etewa S.E., Abdel-Rahman S.A., Fathy G.M., Abo El-Maaty D.A., Sarhan M.H. (2017). Parasitic contamination of commonly consumed fresh vegetables and fruits in some rural areas of Sharkiya government, Egypt. *Afro-Egyptian Journal of Infectious and Endemic Diseases*. 7: 192-202. [DOI: 10.21608/AEJI.2017.17804]
- Fawzi M., El-Sahn A.A., Ibrahim H.F., Shehata A.I. (2004). Vegetable-transmitted parasites among inhabitants of El-Prince, Alexandria and its relation to housewives' knowledge and practices. *The Journal of the Egyptian Public Health Association*. 79: 13-29.
- Hassan A., Farouk H., Abdul-Ghani R. (2012). Parasitological contamination of freshly eaten vegetables collected from local markets in Alexandria, Egypt: a preliminary study. *Food Control*. 26: 500-503. [DOI: 10.1016/j.foodcont.2012.01.033]
- Khan W., Mumtaz G., Bibi S., Afzal S. (2017). Parasitic contamination of fresh vegetables sold at upper and lower Dir Districts, Khyber Pakhtunkhwa, Pakistan. *Pakistan Journal of Zoology*. 49: 1115-1118. [DOI: 10.17582/journal.pjz/2017.49.3.sc3]
- Kudah C., Sovoe S., Baiden F. (2018). Parasitic contamination of commonly consumed vegetables in two markets in Ghana. *Ghana Medical Journal*. 52: 88-93. [DOI: 10.4314/gmj.v52i2.5]
- Marček T., Čorluka S., Gložinić M., Jažić E., Radman P., Sučić M., Ižaković M., Banjari I. (2018). A Comparative survey on the prevalence of parasite elements in fresh vegetables and ready-to-eat salads. *Food in Health and Disease, Scientific-Professional Journal of Nutrition and Dietetics*. 7: 26-30.
- Marques C.S., Sousa S., Castro A., Da Costa J.M.C. (2020). Detection of *Toxoplasma gondii* oocysts in fresh vegetables and berry fruits. *Parasites and Vectors*. 13: 180. [DOI: 10.1186/s13071-020-04040-2]
- Mohamed M.A., Siddig E.E., Elaagip A.H., Edris A.M.M., Nasr A.A. (2016). Parasitic contamination of fresh vegetables sold at central markets in Khartoum state, Sudan. *Annals of Clinical Microbiology and Antimicrobials*. 15: 17. [DOI: 10.1186/s12941-016-0133-5]
- Oladele H.A., Ifeanyi O.E., Akinleye C., Oluwanisola D.O. (2022). Public health risks associated with fruits and vegetables at Owo markets. *International Journal of Current Research in Medical Sciences*. 8: 29-41. [DOI: 10.22192/ijcrms.2022.08.01.003]
- Punsawad C., Phasuk N., Thongtup K., Nagavirochana S., Viriyavejakul P. (2019). Prevalence of parasitic contamination of raw vegetables in Nakhon Si Thammarat province, southern Thailand. *BMC Public Health*. 19: 34. [DOI: 10.1186/s12889-018-6358-9]
- Roberts A.D. (2009). Ascariasis: introduction and epidemiology and transmission. In: Satoskar A.R., Simon G.L., Hotez P.J., Tsuji M. (Editors). Medical parasitology. Landes Bioscience, Austin (Texas). pp: 38-42.
- Said D.E.S. (2012). Detection of parasites in commonly consumed raw vegetables. *Alexandria Journal of Medicine*. 48: 345-352. [DOI: 10.1016/j.ajme.2012.05.005]
- Saida L.A., Nooraldeen K.N. (2014). Prevalence of parasitic stages in six leafy vegetables in markets of Erbil city, Kurdistan region-Iraq. *Zanco Journal of Pure and Applied Sciences*. 26: 25-30.
- Shahnazi M., Jafari-Sabet M. (2010). Prevalence of parasitic contamination of raw vegetables in villages of Qazvin province, Iran. *Foodborne Pathogens and Disease*. 7: 1025-1030. [DOI: 10.1089/fpd.2009.0477]
- Tefera T., Biruksew A., Mekonnen Z., Eshetu T. (2014). Parasitic contamination of fruits and vegetables collected from selected local markets of Jimma town, Southwest Ethiopia. *International Scholarly Research Notices*. 2014. [DOI: 10.1155/2014/382715]
- Traoré S., Bagayoko M.W., Samaké F., Babana A.H. (2021). Assessment of the parasite load of lettuce, tomato and cucumber from some large vegetables production sites in Mali. *African Journal of Agricultural Research*. 17: 1324-1329. [DOI: 10.5897/AJAR2021.15748]
- Vizon K.C.C., Battad Z.G., Castillo D.S.C. (2019). Contamination of food-borne parasites from green-leafy vegetables sold in public markets of San Jose city, Nueva Ecija, Philippines. *Journal of Parasitic Diseases*. 43: 651-657. [DOI: 10.1007/s12639-019-01144-0]
- WHO. (2019). Bench aids for the diagnosis of intestinal parasites. 2nd edition. World Health Organization. URL: <https://www.who.int/publications/i/item/9789241515344>.
- Youssef A.I., Uga S. (2014). Review of parasitic zoonoses in Egypt. *Tropical Medicine and Health*. 42: 3-14. [DOI: 10.2149/tmh.2013-23]